



Wildlife poop too: investigating the role of introduced predators and avian species on microbial water quality assessments

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agresearch
āta mātai, mātai whetū

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Science for Communities

E. coli = faecal indicator bacteria

- Diverse sources of water pollution

But...

lack of data on wildlife

- Wildlife shown to harbour “cryptic *Escherichia* clades” in their faeces (e.g. *E. marmotae*, *E. ruysiae*)
- Routine “*E. coli*” monitoring tests cannot distinguish faecal *E. coli* from those “cryptic clades”



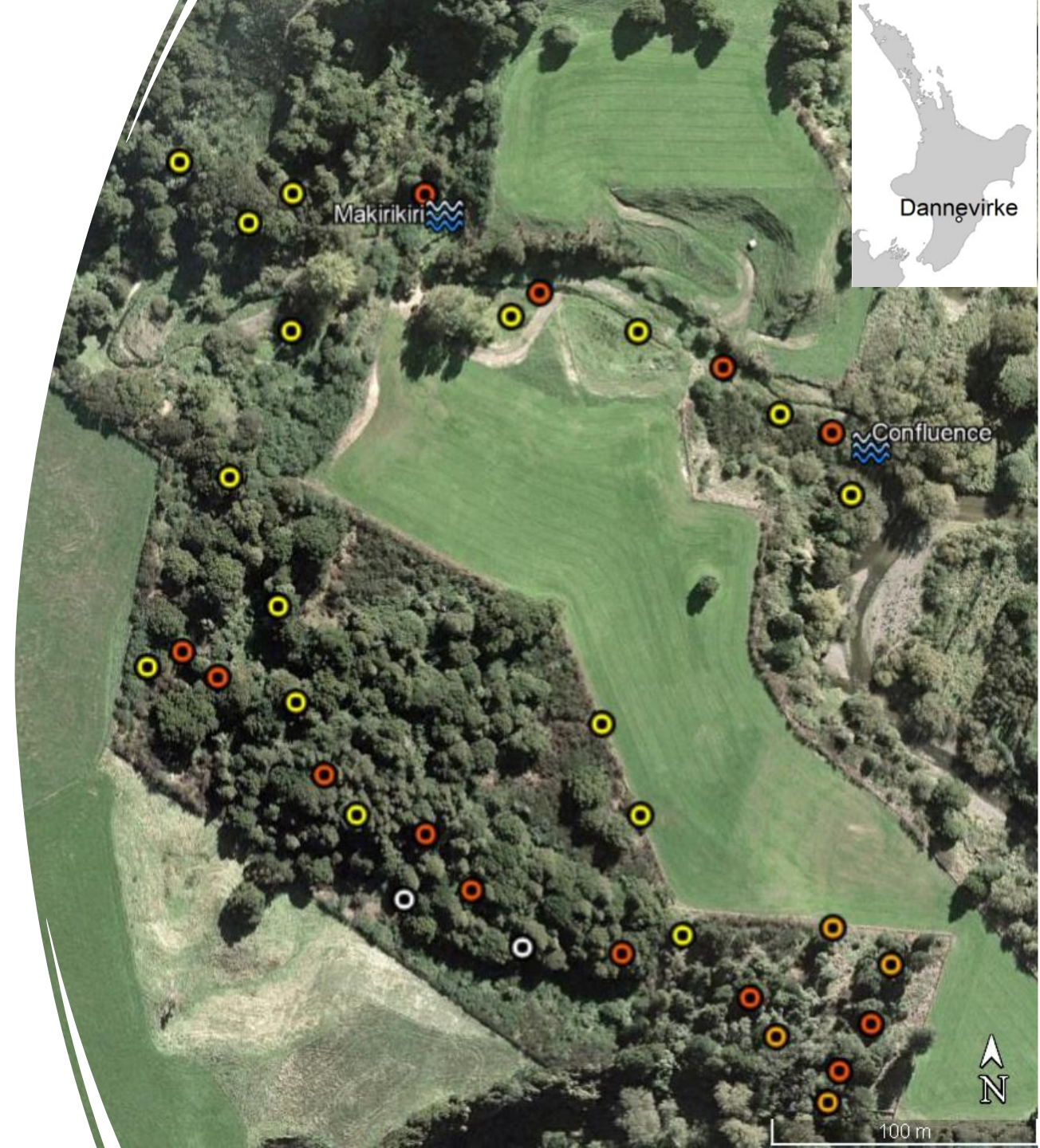
Aims

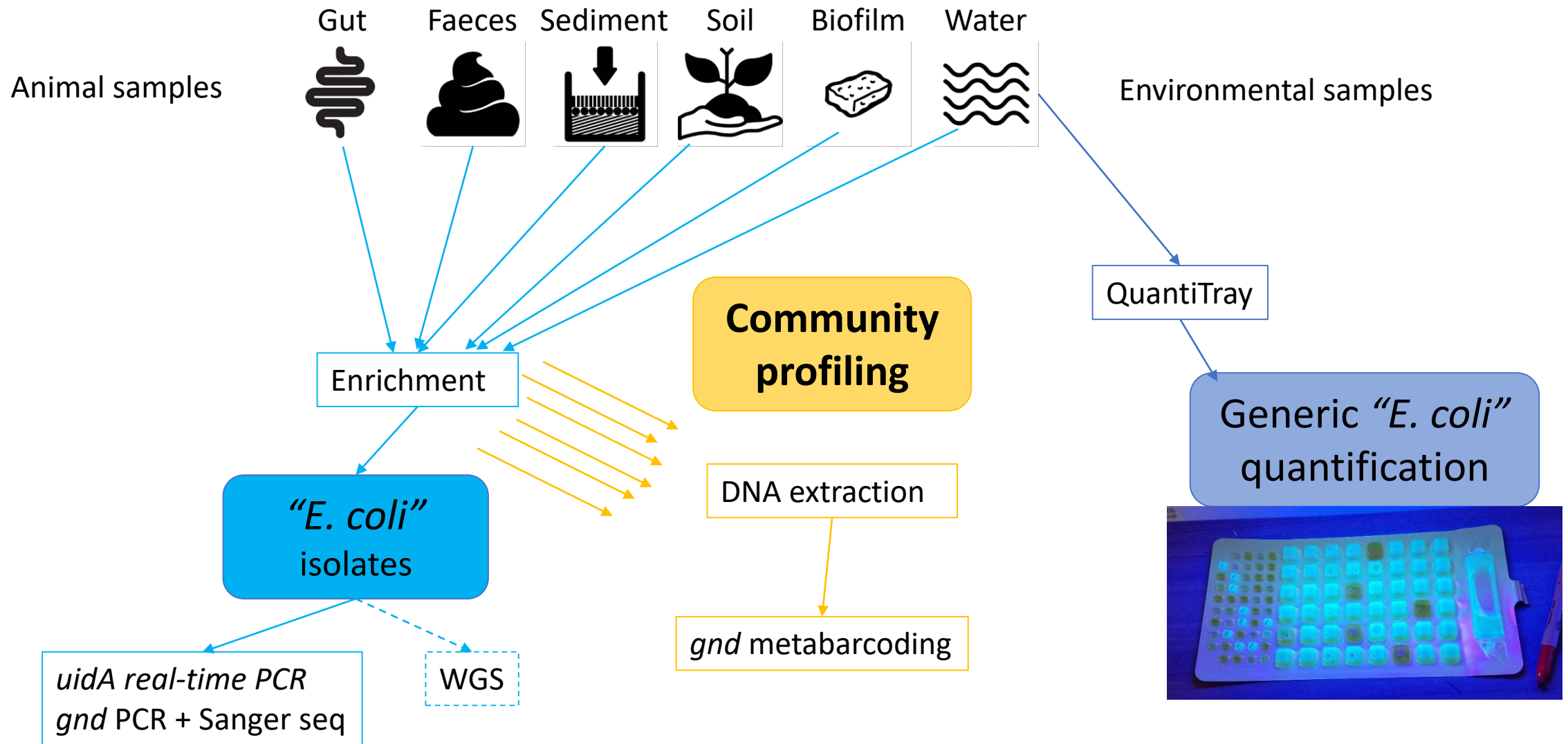
- Co-design and co-innovation of project undertaken with local hapū
- Provide opportunities for local hapū to reconnect with culturally significant Mākirikiri Reserve
- Support hapū with tools to undertake pest management to enhance the mauri of the ngahere during project and after
- Examine faecal material from bird and introduced predator species
 - WGS and phylogenomic analysis of “*E. coli*” isolates
 - “*E. coli*” community profiling (*gnd* metabarcoding)
- Compare with environmental samples



Methods

- Mākirikiri Reserve/Stream, Dannevirke
- Animal trapping (Nov-Dec 2020)
 - 13x GoodNature A24 
 - 4x GoodNature A12
 - 16x Timms traps 
 - 2x Trapinator
- Environmental sampling (4 visits)
 -  Water, soil, sediment, periphyton
 - 2 sites
- Faeces from environment
 - Avian
 - Mammal





OPEN

Culture independent analysis using *gnd* as a target gene to assess *Escherichia coli* diversity and community structure





1: 17 November 2016
 1: 16 March 2017
 1 online: 12 April 2017

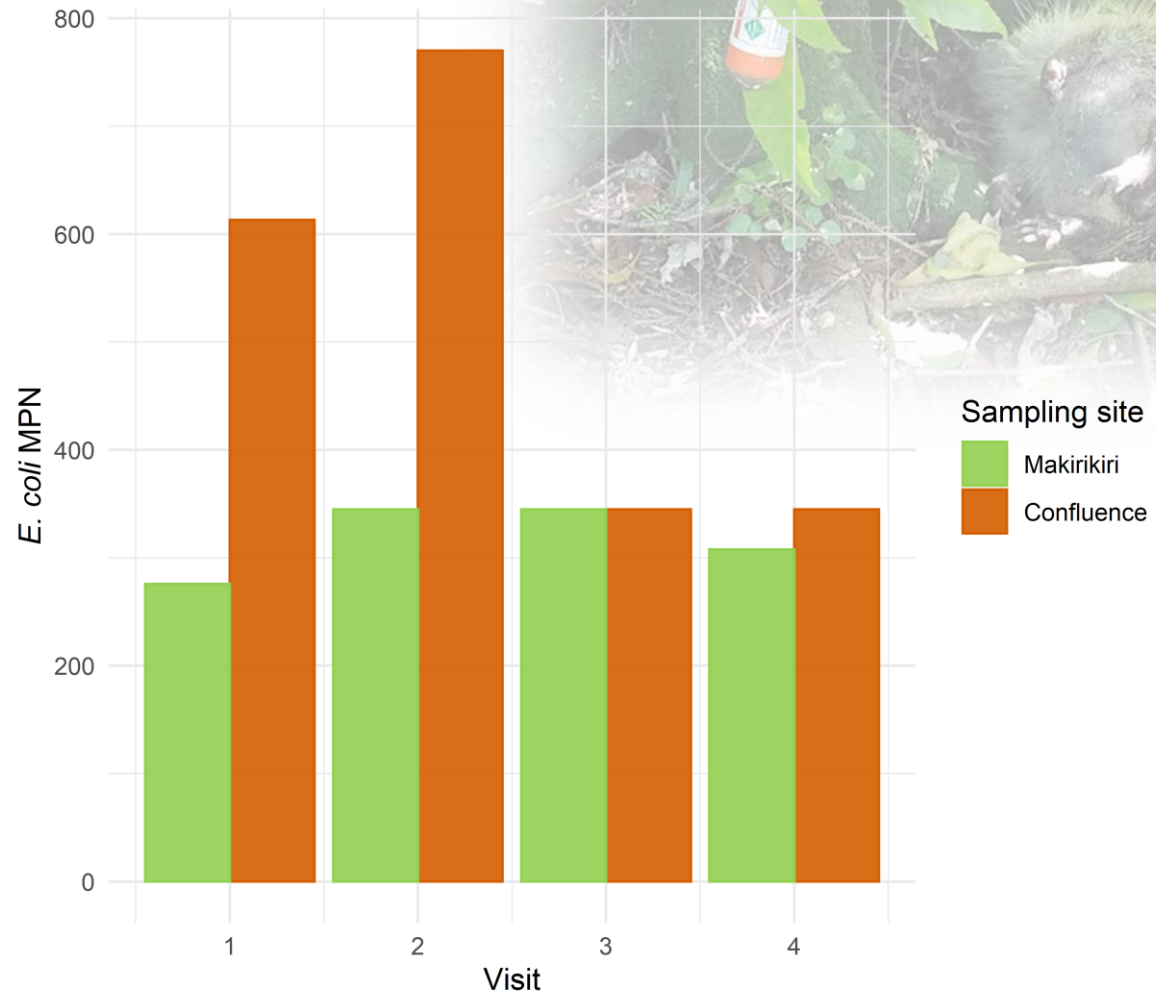
Adrian L. Cookson^{1,2}, Patrick J. Biggs^{2,3}, Jonathan C. Marshall^{2,4}, Angela Reynolds¹, Rose M. Collis¹, Nigel P. French² & Gale Brightwell¹

***gnd*Db, a Database of Partial *gnd* Sequences To Assist with Analysis of *Escherichia coli* Communities Using High-Throughput Sequencing**

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Results

- 1523.5 corrected trap-nights
 - 49  5 
 - 3  1 
- 58 Gut samples
+ 16 faecal samples
(7 mammal/ 9 avian)
- 32 environmental samples
- 420 isolates
 - 207 underwent *gnd* sequence typing (gST)
 - 102 sent for WGS



Diversity of isolates

- *uidA* gene not amplified in 32/420 isolates (12 samples)

4  5  2  1 avian 

- Animal (gut + faeces):
36 gSTs/143 isolates

- Environment:
41gSTs/64 isolates

- gST258 and gST535 most frequent gSTs (almost ½ of animal samples)



- gST258

- 1 sediment 

- 3 avian 


- 22  +1 

- +1  +1 

- gST535

- 2 water 

- 1 soil 

- 3 mammal  +1 
- 17 

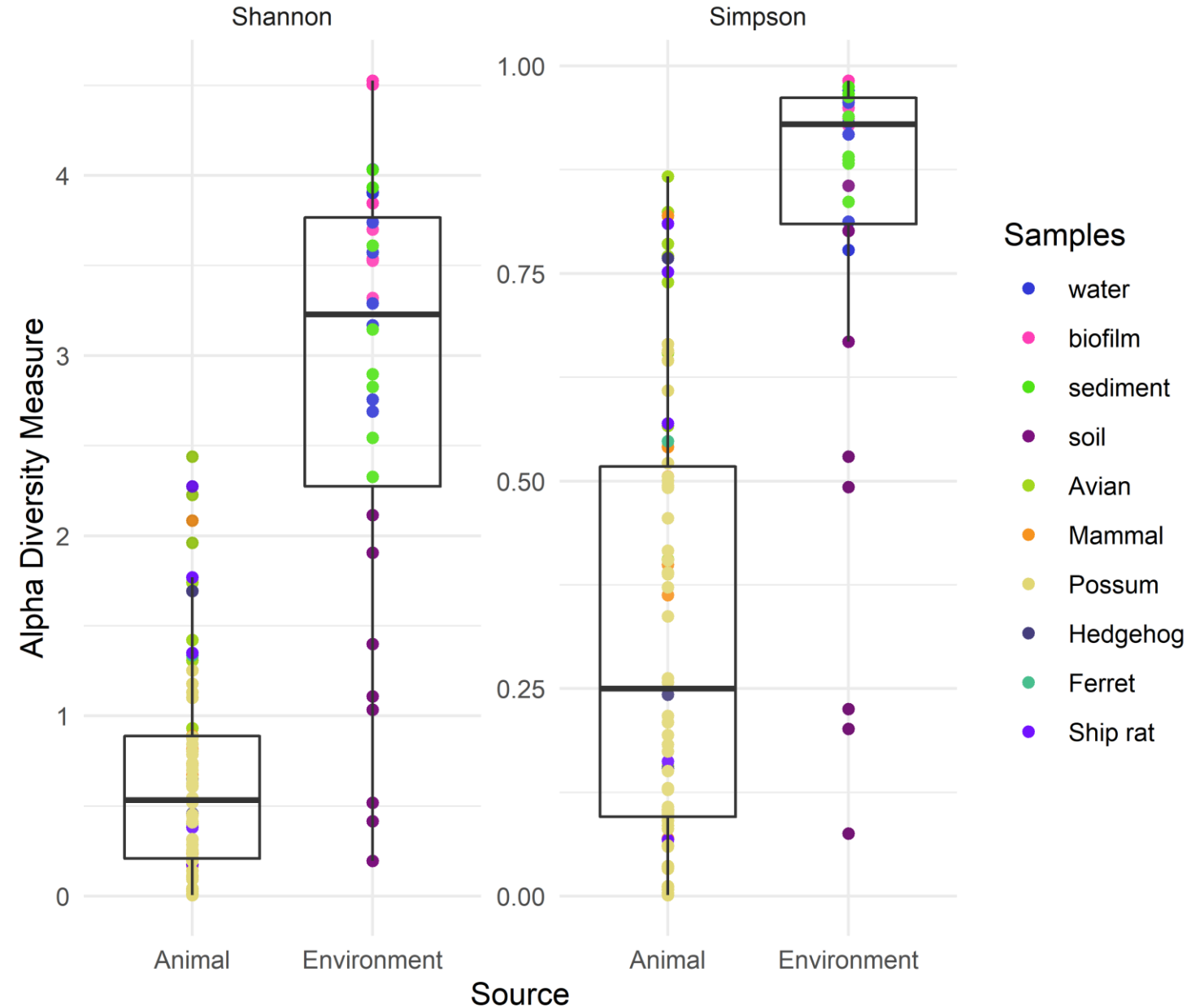
Community composition (*gnd* metabarcoding)

SAMPLE TYPE	NO.	NUMBER OF <i>g</i> STs DETECTED	% OF TOTAL	MEAN NUMBER OF <i>g</i> STs PER SAMPLE +/- SD
Water	8	267	52%	87.5 +/- 14
Sediment	8	207	40%	55.6 +/- 31
Soil	8	84	16%	13.5 +/- 6.5
Biofilm	8	301	58%	131.5 +/- 34.9
Faeces	16	196	38%	16.2 +/- 11.3
Avian	9	131	25%	21.4 +/- 10.3
Mammal	7	45	9%	9.4 +/- 9.1
Gut content	58	173	34%	7.1 +/- 8.8

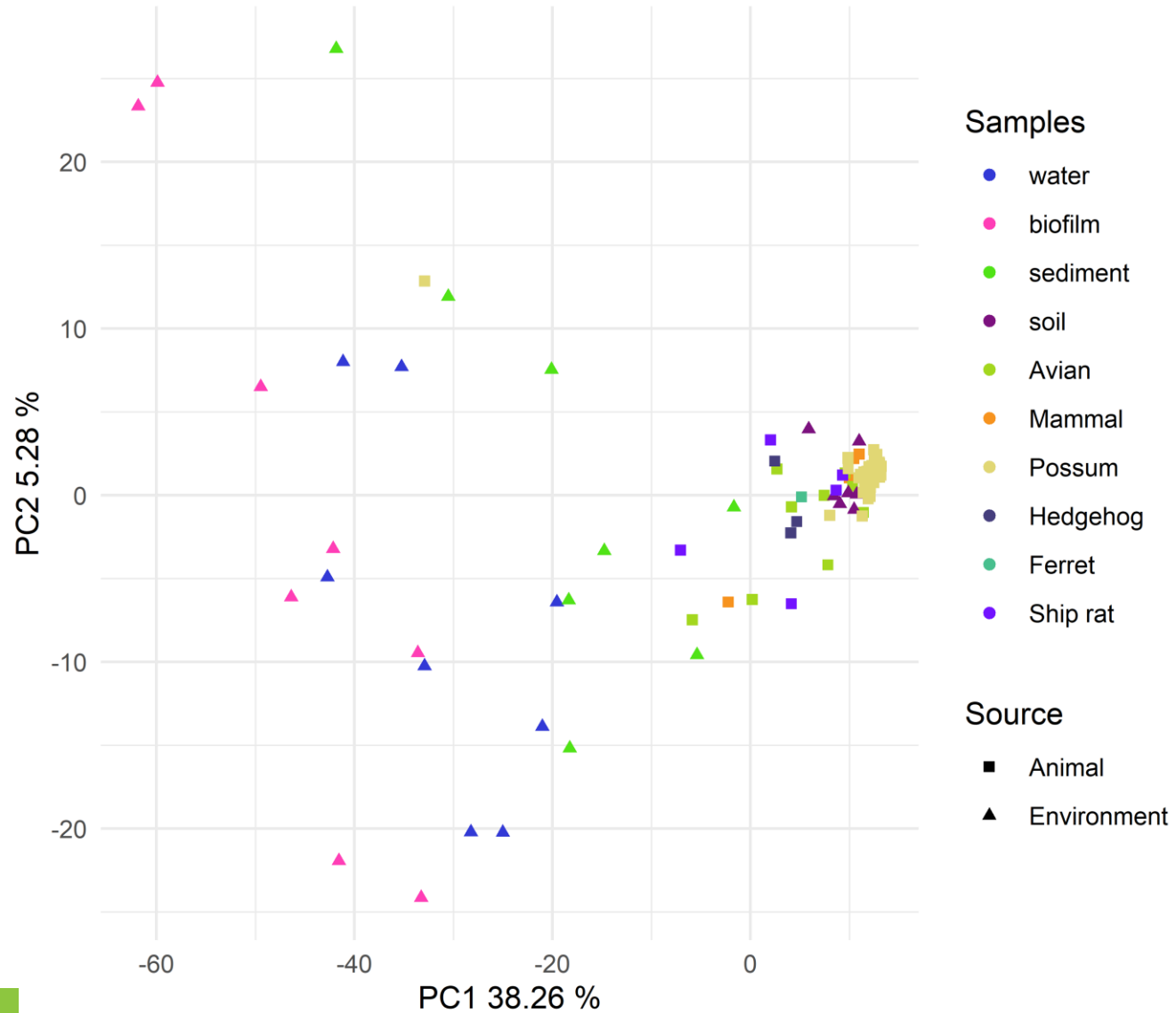
Alpha diversity of gSTs

Significantly lower for
animal samples

P-value << 0.001



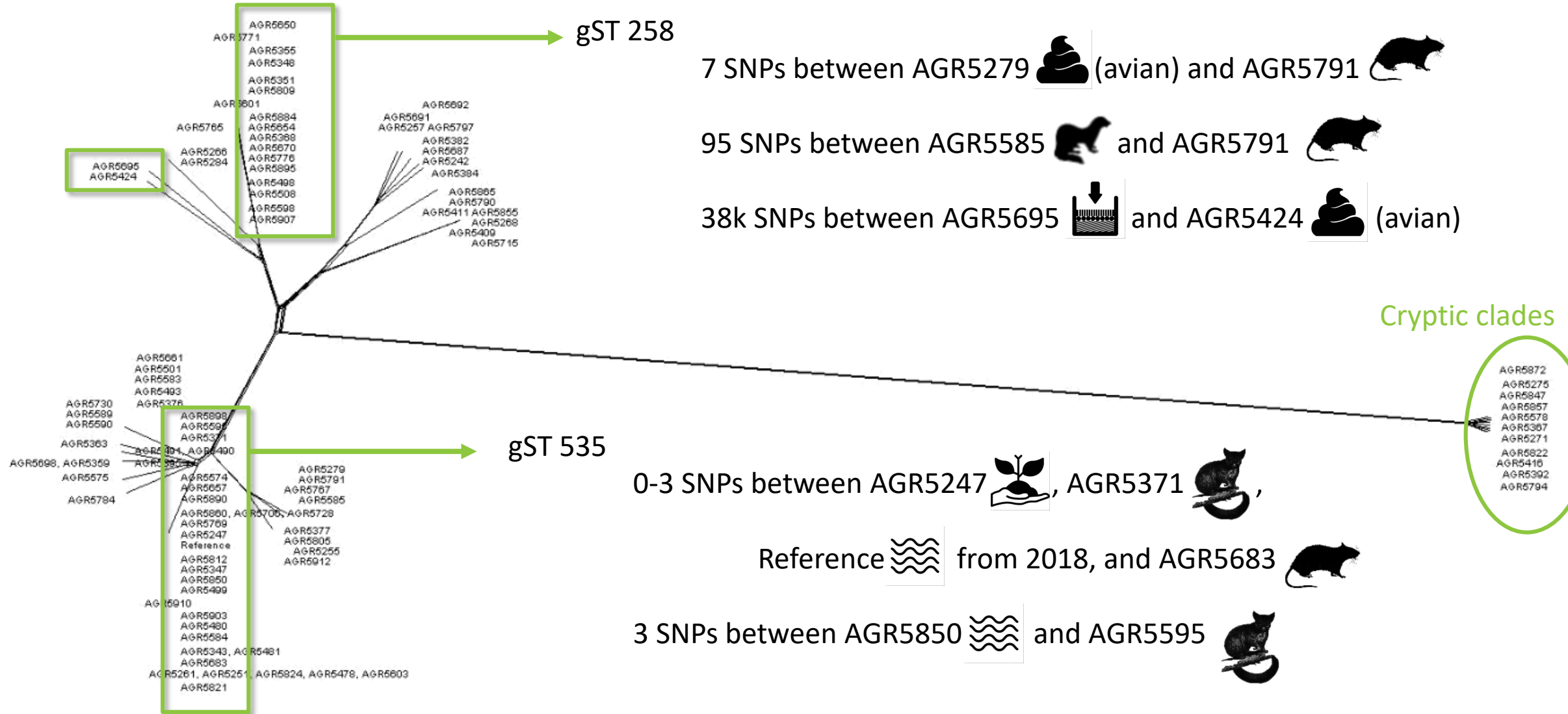
PCA



Most frequent gSTs (with ≥ 10 reads)

Most frequent gSTs	in Animal samples	in Environmental samples	Overall
gST535	62 (84%)	28 (88%)	90 (85%)
gST258	55 (74%)	20 (63%)	75 (71%)
gST522	12 (16%)	26 (81%)	38 (36%)
gST152	11 (15%)	22 (69%)	33 (31%)
gST514	11 (15%)	22 (69%)	33 (31%)
gST308	14 (19%)	17 (53%)	31 (29%)
gST587	13 (18%)	15 (47%)	28 (26%)
gST231	5 (7%)	22 (69%)	27 (25%)

WGS analysis



→ Stable and long-lasting clone in the environment

Conclusions

- gSTs shared between animal and environmental samples
 - Wildlife = source (among others) of environmental contamination
 - Cryptic clades found only in low abundance
 - Traps were gifted to local hapū at the end of the project to continue trapping
- Pest management could conceivably improve water quality (as well as improve biodiversity)**



Acknowledgements

- Co-authors



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- Te Kāuru (Eastern Manawatū River) hapū collective and other local hapū
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- Lauren Gadd - Pūhoro STEMM Academy intern student
- Rose and Tracey Collis for carcass logistics



Questions?

